# Reference Values of Lymphocyte Sub-Populations in Healthy Human Immunodeficiency Virus-Negative Iranian Adults

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### ABSTRACT

**Background:** Lymphocyte subsets enumeration is considered prominent in the management of primary and acquired immunodeficiency disorders. Because of local variations due to race, age, gender, and environmental conditions on lymphocyte subsets, and to improve the accuracy of interpretation of laboratory findings, reference intervals must be determined in every population. Objective: To establish a normal reference range for CD3<sup>+</sup>, CD4<sup>+</sup>, CD8<sup>+</sup>, CD19<sup>+</sup> and CD56<sup>+</sup> lymphocytes in a healthy Iranian adult population using flowcytometry. Method: Blood samples were collected from 221 HIV seronegative individuals, including 112 females and 109 males, with ages ranging from 20 to 40 years old. The percentage of lymphocytes expressing either of CD3, CD4, CD8, CD19 and CD56 surface markers were determined by flowcytometry assay. Result: Total mean percentage and absolute count of lymphocyte subsets were as follows:  $CD3^+$ : 70.90 ± 7.54%, 1800.87 ± 471.09 cells/µl;  $CD4^+$ : 41.04 ± 7.86%,  $1039.99 \pm 338.02$  cells/µl; CD8<sup>+</sup>:  $31.11 \pm 6.60\%$ ,  $783.95 \pm 234.87$  cells/µl; CD19<sup>+</sup>:  $12.77 \pm 4.56\%$ ,  $328.37 \pm 153.17$  cells/µl; CD56<sup>+</sup>:  $15.53 \pm 6.34\%$ ,  $388.62 \pm 176.17$ cells/ $\mu$ l, respectively. The ratio of CD4<sup>+</sup>/CD8<sup>+</sup> lymphocytes for the studied population was  $1.39 \pm 0.48$ . Significant differences were observed between male and female subjects indicating that the average percentage of  $CD3^+$  cells (p=0.017) and  $CD4^+$  T cells (p=0.003) were higher in the female population, whereas the average percentage of  $CD19^+$  cells (p=0.02) tended to be higher among males. However, investigations on the CD56<sup>+</sup> NK cell and CD8<sup>+</sup> T cell sub-populations did not show any statistical differences between the two genders. In comparison with reports of other populations, we were confronted with different results. Conclusion: Establishing reference values of lymphocyte subsets for each population is helpful in achieving standard criteria for the prognosis of HIV infection. Therefore, normal ranges established by this survey can be used as a reference for decisions made in clinical practice.

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Keywords: HIV Seronegativity, Immunophenotyping, Lymphocyte Subsets, Reference Value

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### INTRODUCTION

Immunohematological parameters are affected by several factors including genetics, age, gender, environment (e.g. nutrition, drug intake, smoking, dietary patterns), immunological disorders (such as autoimmune disorders, infection, lymphoma, leukemia, immunodeficiency) and so on. As the differentiation of these factors depends on the population and geographical region studied, racial differences and variations of immunohematological parameters exist worldwide. On the other hand, ethnic and racial differences are remarkable elements which influence the level of immune cells such as lymphocyte subsets. With this in mind, it is considerable that these parameters could be used as alternative markers for the assessments of immune status in various diseases (1). In this context, the notable example is Human Immunodeficiency Virus syndrome (HIV). The stated parameters could be used for the management of diagnosis and disease progression as well as decisions regarding prophylactic therapies against opportunistic infections and response to antiretroviral treatment in HIV infected patients (2). Attempting to understand the pathogenesis of HIV infection has prompted careful study of immune cells, demonstrating that the prominent changes in lymphocyte subpopulations including T cells, B cells and NK cells are associated with HIV infection (3-6). These lymphocyte subpopulations are distinguishable by their surface CD markers (clusters of differentiation) (7). B cells are defined by the expression of either CD19 or CD20; Natural Killer (NK) cells are determined by the presence of CD16 or/and CD56 and negativity for CD3; T cells are characterized as the cells expressing CD3 antigen and T cell subpopulations (Helper, Suppressor/Inducer) are identified by the coexpression of CD4 or CD8. With this background, it is beneficial to develop reference values of immunohematological parameters for each population individually in order to improve the quality of health care. Currently, flowcytometry is considered as the standard technology for quantification of lymphocyte subsets. Thus, lymphocyte immunophenotyping classified on the basis of their CD markers could be used as an important tool for the evaluation of immunological status (8). Therefore, this cross sectional study was performed to define normal reference ranges of both percentage and absolute count of peripheral blood lymphocyte subsets in HIV seronegative Iranian males and females with age of 20 to 40 years using flowcytometry. With regard to immunodeficiency syndromes such as HIV, the present reference ranges could be advantageous for diagnosis, evaluation of disease stage and progression, and therapeutic monitoring in Iranian individuals with HIV infection. Finally, we compared our results with other similar studies in order to point out the disparities of normal values of lymphocyte subsets in different populations.

## MATERIALS AND METHODS

**Subjects.** Between December 2010 and December 2012, a total of 221 medical students of Tehran University of Medical Sciences and University-affiliated hospital employees were recruited to this study. Participants, who were comprised of 109 males and 112 females, were grouped into four age categories: 20-24 years, 25-29 years, 30-34 years and 35-40 years. HIV testing was done for all consenting individuals using the same kits and procedures. Subsequently, subjects who were seronegative for HIV-1 and had no history of smoking, alcohol consumption, and drug abuse were involved in our study.

The following exclusion criteria were considered: any sickness within the last month (including viral infection or parasitic infection); any chronic disease; any vaccination (within the last 4 weeks); any major surgery (within the last 6 months); pregnancy/ lactation (within the last 6 months). The characteristics of individuals are listed in Table 1. The study was approved by the Ethics Committee of Tehran University of Medical Sciences and its related hospitals and all subjects gave an informed written consent prior to enter this study.

**Sample Collection and HIV Serology.** Two blood samples were drawn from each participant. One specimen was collected in a 5 ml anticoagulant-free tube for HIV testing and the second one in a 5 ml EDTA-containing tube for determination of hematological indices. In order to determine the seronegativity for HIV, serum samples were analyzed using enzyme-linked immunosorbent assay (Virinostika HIV Ag/Ab ELISA, BioMerieux, France) based on a one-step sandwich test. Specific Microelisa wells were coated with HIV-1 gp160, HIV-1 ANT70 peptide, HIV-2 env peptide (amino acids 592-603) and anti-HIV-1 p24. All samples were analyzed on the day of collection.

**Hematological Analysis.** A Hemacytometer (Sysmex KX21, Japan) device was used for the whole-blood analysis of hematological parameters. After automatic dilution of the whole-blood sample, lysis, and count the device gave a printout result of absolute numbers of leukocytes (WBC), platelets, neutrophils, eosinophils, monocytes, Bands (all expressed as number of cells\* $[10^9]$ /lit), lymphocytes (expressed as number of cells\* $[10^9]$ /lit), lymphocytes (gram per deciliter). Besides, ESR (mm/hour) was reported using the Westergren method.

Flowcytometric Analysis. Lymphocyte cells were analyzed using a FACSCalibur Flowcytometer (Becton Dickinson Immunocytometry Systems). Specimens were tested to determine the proportion of T lymphocytes identified by the expression of CD3, Tcell subpopulations (T Helper Cells and Cytotoxic T Cells) by the co-expression of CD4 or CD8, B lymphocytes by the expression of CD19 and Natural Killer (NK) lymphocytes by the positivity of CD16 and/or CD56 markers and lack of CD3 expression. This was performed by incubating anticoagulated whole blood with monoclonal antibodies (MAb) which were purchased from Becton Dickinson: CD3-FITC/CD19PE, CD4-FITC/CD8-PE and CD56-PE. Briefly; 100µl of whole blood was mixed with 10µl of the appropriate MAb combination and then incubated at room temperature for 20 mins in the dark. RBCs were then lysed by adding 1 ml of fluorescence activated cell sorter lysing solution (Becton Dickinson). After vortexing, tubes were incubated in the dark at room temperature for 10 mins and then centrifuged at 252 RCF of G for 5 min. The cell pellet was washed once with 1 ml of Facs Flow<sup>®</sup>, resuspended in 500 µl of Facs Flow, on forward and side scatter the lymphocytes were acquired, gated and then analyzed (Figure 1) using the Cell Quest Pro software (Becton Dickinson). Using cell blood count, absolute values of lymphocyte subsets were obtained.

**Statistical Analysis.** Statistical analysis was performed using SPSS software (version 14.0). One-sample Kolmogorov-Smirnov test was performed to find out whether the variables were normally distributed or not. Since the T cell subsets had a normal distribution, the T cell subset proportions and hematological parameters were compared between males and females using Student's *t-test*. The age-related differences were also analyzed by ANOVA test. For each immunohematological parameter the average value

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and standard deviation (SD) was calculated and expressed as the mean  $\pm$  SD. Results with a p value less than 0.05 were considered statistically significant.



Figure 1. Example of CD3, CD4, CD8, CD19 and CD56 analysis FITC-RPE negative control.

### RESULTS

**Characteristics of Studied Subjects.** Blood samples from a total of 221 healthy Iranians, including 109 males (49.3%) and 112 females, (50.7%) were collected and screened for HIV infection. Afterwards, the effect of age and gender of the HIV seronegative participants on immunohematological indices was survived by categorizing each gender group into four age subgroups including 20-24 years, 25-29 years, 30-34 years and 35-40 years. Using immunohemotyping, specimens were tested for the proportion of lymphocyte subsets comprised of CD3<sup>+</sup> T cells, sub-populations of T cells (CD4<sup>+</sup> T helper and CD8<sup>+</sup> T suppressor/inducer), CD19<sup>+</sup> B cells and CD56<sup>+</sup> NK cells, considering sex and age group. Distribution in each age group according to gender is presented in Table 1. The mean age range for females and males was 29.07  $\pm$  6.1 and 28.74  $\pm$  6.1, respectively.

Gender	Male			Female	Total		
Age group	No.	Mean ± SD	No. Mean ±		No.	Mean ± SD	
20-24 years	32	$22.28 \pm 1.67$	33	$21.87 \pm 1.49$	65	22.07 ± 1.58	
25-29 years	28	$26.85 \pm 1.62$	29	$25.55\pm0.68$	57	26.19 ± 1.39	
30-34 years	27	$31.59 \pm 1.27$	26	$30.84 \pm 1.25$	53	$31.22 \pm 1.31$	
35-40 years	22	$38.68 \pm 1.61$	24	$38.25\pm2.0$	46	$38.45 \pm 1.84$	
Total	109	$29.07\pm6.1$	112	$28.41\pm6.2$	221	$28.74 \pm 6.1$	

Table 1. Age distribution of subjects for both genders according to age groups.

**Hematological Analysis in HIV-Seronegative Iranian Subjects.** The results of the hematological parameters according to gender were as follows: Platelets:  $242.83 \pm 52.79$  (females) and  $218.92 \pm 43.01$  cells/µl (males); Neutrophils:  $3588.79 \pm 971.07$  (females) and  $3619.18 \pm 956.87$  (males) cells/µl; Monocytes:  $291.098214 \pm 93.417494$  (females) and  $313.899083 \pm 108.195805$ ; Band:  $44.348214 \pm 61.918387$  (females) and  $54.541284 \pm 78.765655$  (males); Eosinophil:  $136.696429 \pm 130.089771$  (females) and  $172.862385 \pm 120.142159$  (males); hemoglobin:  $13.10 \pm 1.12$  (females) and  $14.68 \pm 1.31$  (males) g/dl; ESR:  $8.16 \pm 5.35$  (females) and  $5.66 \pm 3.25$  (males) mm/hr. According to this gender-related data it was confirmed that the average Platelet count (p=0.02) and ESR (p=0.01) were higher in female subjects, whereas the average count for Eosinophil (p=0.03) and hemoglobin level (p=0.05) were statistically higher in males. None of the other studied hematological indices represented a significant difference between the two genders (p>0.05).

The mean value of total lymphocyte count among different age groups is given in Table 2 As a result, no gender-specific difference was observed for total lymphocyte enumeration (p>0.05). Moreover, no apparent age-dependent differences for lymphocyte subpopulation were noted in the data.

Lymphocyte count								
Gender	Female							
Age Group	Mean $\pm$ SD	Mean $\pm$ SD	P-value					
20-24 years	$2530.06 \pm 663.45$	$2493.71 \pm 622.46$	NS					
25-29 years	$2414.00 \pm 615.79$	$2572.64 \pm 599.54$	NS					
30-34 years	$2660.11 \pm 572.90$	$2508.29 \pm 529.92$	NS					
35-40 years	$2482.87 \pm 617.49$	$2694.22 \pm 623.48$	NS					
Total	$2520.08 \pm 619.14$	$2558.07 \pm 591.56$	NS					

Table 2. The mean  $\pm$  SD of lymphocyte counts for both genders among age groups.

NS: non-significant

**Immunophenotyping of Lymphocyte Sub-Populations in HIV-Negative Iranian Subjects.** Table 3 displays the average of absolute counts and percentages of lymphocyte subsets in 112 female and 109 male subjects. Regarding gender-specific comparison, the mean percentages of  $CD3^+$  T cells and  $CD4^+$  T cells were significantly higher in females but not supported by absolute values, whereas the mean percentage of  $CD19^+$  B cells tended to be higher in males.

Gender		Female (112)	Male (109)		Total	
Cell subsets		Mean $\pm$ SD	Mean $\pm$ SD	P-value	Mean $\pm$ SD	
CD3 <sup>+</sup> T cells	Absolute value	1816.73 ±482.21	$1784.57 \pm 461.4$	NS	$1800.87 \pm 471.09$	
	%	$72.09 \pm 7.54$	$69.68 \pm 7.37$	0.017*	$70.90\pm7.54$	
CD4 <sup>+</sup> T cells	Absolute value	$1072.05 \pm 384.90$	$1007.05 \pm 279.85$	NS	$1039.99 \pm 338.02$	
	%	$42.56 \pm 8.69$	$39.48\pm 6.59$	0.003*	$41.04\pm7.86$	
$CD8^+$ T cells	Absolute value	$777.42 \pm 223.77$	$790.67 \pm 246.61$	NS	$783.95\pm234.87$	
	%	$31.20 \pm 6.51$	$31.01\pm6.72$	NS	$31.11 \pm 6.60$	
CD19 <sup>+</sup> B cells	Absolute value	309.91 ± 143.99	$347.33 \pm 160.51$	NS	$328.37 \pm 153.17$	
	%	$12.11 \pm 3.89$	$13.45\pm5.08$	0.02*	$12.77\pm4.56$	
CD56 <sup>+</sup> NK cells	Absolute value	387.57 ± 183.30	389.69 ± 169.37	NS	$388.62 \pm 176.17$	
CD4 <sup>+</sup> /CD8 <sup>+</sup> ratio	%	$15.64 \pm 6.80$ $1.44 \pm 0.53$	$\begin{array}{c} 15.42 \pm 5.87 \\ 1.35 \pm 0.42 \end{array}$	NS NS	$\begin{array}{c} 15.53 \pm 6.34 \\ 1.39 \pm 0.48 \end{array}$	

# Table 3. The mean $\pm$ SD percentages and absolute values of lymphocyte subsets in 112 female and 109 male healthy adults.

NS:non-significant

	<b>20-24</b> years		25-29 years		30-34	4 years	35-40 years	
	Female	Male	Female	Male	Female	Male	Female	Male
	Mean ±SD	Mean ±SD						
CD3 <sup>#</sup>	$1805.92 \pm 547.83$	1706.16 ± 455.24	1763.12 ± 438.74	1814.50 ± 466.42	1890.22 ± 485.36	1768.34± 431.34	1816.75 ± 452.03	$1880.46 \pm 506.60$
%	$70.96 \pm 7.12$	$68.62\pm8.38$	$73.31 \pm 5.66$	$70.48\pm 6.92$	$70.90\pm9.96$	$70.17 \pm 6.15$	$73.47\pm7.02$	$69.63\pm8.02$
CD4 <sup>#</sup>	$1000.24 \pm 450.96$	$999.50 \pm 304.34$	$1086.36 \pm 315.36$	967.51 ± 259.14	$1160.91 \pm 349.86$	$990.95 \pm 286.07$	$1057.21 \pm 401.37$	$1088.12 \pm 262.68$
%	$40.07\pm8.71$	$39.98 \pm 6.14$	45.32* ± 8.45	$37.69 \pm 5.35$	$43.14 \pm 7.46$	$39.36 \pm 7.07$	$42.01\pm9.62$	41.16 ± 7.82
$\text{CD8}^{\#}$	$826.75 \pm 230.50$	$744.43 \pm 220.80$	$755.30 \pm 199.12$	$858.36 \pm 266.79$	$754.80\pm233.19$	$803.57 \pm 238.23$	$760.80 \pm 235.35$	755.94 ± 261.11
%	33.07*± 5.67	$29.98\pm 6.03$	$31.90 \pm 6.72$	33.48 ± 6.63	$28.49\pm6.7$	$32.05 \pm 6.83$	$30.73 \pm 6.48$	$28.07 \pm 6.64$
CD19 <sup>#</sup>	$311.19 \pm 126.98$	$354.04 \pm 182.69$	$315.98 \pm 161.60$	$353.26 \pm 145.18$	$328.30 \pm 159.14$	$315.09 \pm 158.50$	$280.92* \pm 130.39$	369.59 ± 151.83
%	$12.18 \pm 3.23$	$13.97 \pm 5.6$	$12.67 \pm 4.13$	$13.72 \pm 4.96$	$12.26 \pm 4.51$	12.31 ± 5.23	11.17* ± 3.79	$13.75 \pm 4.32$
CD56 <sup>#</sup>	$429.29 \pm 146.56$	359.09 ± 175.08	340.52 ± 132.37	422.42 ± 178.42	375.90 ± 197.96	$406.04 \pm 124.17$	399.68 ± 250.49	372.49 ± 196.85
%	17.58* ± 5.79	$14.41 \pm 5.77$	$14.56\pm5.92$	16.62 ±5.95	$14.30 \pm 7.58$	$16.60 \pm 5.42$	$15.71 \pm 7.92$	$13.93 \pm 6.2$
CD4/CD8 ratio	$1.22 \pm 0.45$	$1.38 \pm 0.34$	$1.51^* \pm 0.55$	1.19 ± 0.39	1.63* ± 0.61	$1.31 \pm 0.46$	$1.43 \pm 0.44$	$1.55 \pm 0.47$

 Table 4. The mean ± SD percentages and absolute values of lymphocyte subsets according to age groups.

\*: P-value less than 0.05

#: Absolute value,

However, no significant differences in the average percentages and absolute counts of other lymphocyte subsets (CD8<sup>+</sup> T cells and NK cells) were observed (p>0.05). Similarly, no statistically significant difference was noted for the mean value of CD4/CD8 ratio between the two genders (p>0.05).

Table 4 represents a comparison between the average percentages and absolute values of lymphocyte sub-populations for each studied age group. According to age-group comparisons between the two genders, some parameters were higher in female subjects including: the average percentage of  $CD8^+$  T cells (p=0.037) and  $CD56^+$  NK cells (p=0.031) in 20-24 years age group; mean percentage of  $CD4^+$  T cells (p=0.00) in 25-29 years age group. Absolute value and average percentage of  $CD19^+$  B cells (p=0.04) in 35-40 years age group was higher in males in comparison to females. In addition, the mean value of  $CD4^+/CD8^+$  ratio in the 25-29 years (p=0.015) and 30-34 years (p=0.037) age groups was higher in females compared to males. According to the population's age classification, our results did not show any significant variation of lymphocyte subsets among the considered age groups.

## DISCUSSION

This study aimed to establish age and gender specific immunohematological reference values which may be considered as the Iranian standard for the follow-up of HIV-positive patients. Generally, analysis of lymphocyte subsets is a crucial aspect in the management of patients with HIV. With more details, the importance of establishing absolute counts and percentage values of lymphocyte subsets and, especially, the ratio of  $CD4^+/CD8^+$  T cells is considerable in manipulating the progression of HIV infection, guiding the initiation of treatment, and monitoring responses to antiviral therapy. Region-specific reference values of peripheral lymphocyte subtypes have been determined by several studies throughout the world and have exhibited some discrepancies based on geographical region and methodology. Additionally, it is suggested that these variations could also be attributed to ethnical diversity, genetics, environmental factors such as nutrition, climate, smoking, exposure to different antigens and methods of enumeration. The accurate interpretation of immunophenotyping data, including T cell sub-populations' counts, is reliant on age-specific lymphocyte subsets' reference values of healthy subjects in the relevant population (9,10,11).

Therefore, we have enumerated lymphocyte subsets in our population using venous blood obtained from 221 healthy HIV-negative volunteers stratified by age, ranging from 20 to 40 years. It is evident that the results are not creditable for children and the elderly. Immunophenotyping by flowcytometry was used for the evaluation of lymphocyte subsets including T, B and NK cells.

Based on our hematological results, it was shown that both average platelet count (p=0.00) and rate of ESR (p=0.00) were increased in females. Nevertheless, the average eosinophil count (p=0.03) and hemoglobin level (p=0.00) were statistically higher in males. Besides, with respect to age-dependent changes, differences among groups were not worthy of reflection. Similarly, no age-related differences between males and females were identified for total lymphocyte count among the four studied age groups (p>0.05).

With respect to gender-specific differences in lymphocyte subsets, Iranian females were found to have significantly higher percentages of CD3<sup>+</sup> T and CD4<sup>+</sup> T cells, whereas the

percentage of CD19<sup>+</sup> B cells was higher in males. However, comparison of absolute values failed to show any statistically significant difference for the mentioned lymphocyte subsets (p>0.05). The mean percentage and absolute values of NK cells and CD8<sup>+</sup> T cell subsets were also found to be gender-independent (p>0.05). We observed no differences in the mean CD4<sup>+</sup>/CD8<sup>+</sup> ratio between males and females (p>0.05). One possible explanation for the observed gender differences in the average percentage of CD4<sup>+</sup>, CD3<sup>+</sup> and CD19<sup>+</sup> cells, is that it may be secondary to the differential effects of sex hormones (12,13).

Considering the studied age groups, we identified the average percentage and absolute count of each sub-type among four groups in each sex. As a summary, in the 20-24 years age group: the average percentage of  $CD8^+$  T cells (p=0.037) and the average percentage of  $CD56^+$  NK cells (p=0.031); in the 25-29 years age group: the average percentage of  $CD4^+$  T cells (p=0.00) were higher in females than males and in the 35-40 years age group: the absolute value and average percentage of  $CD4^+$  CD8<sup>+</sup> ratio, the so-called helper/suppressor ratio, for the 25-29 years (p=0.015) and the 30-34 years age groups (p=0.037) was higher in females compared to males.

As a whole, our investigated immunohematological parameters did not increase or decrease in ages between 20 to 40. The influence of age on lymphocyte sub-populations is not yet well established, since different studies report different results; various studies have illustrated that the number of  $CD4^+$  T cells increases, whereas the number of  $CD8^+$  T cells decreases with age (13), while others have demonstrated that the number of both  $CD4^+$  and  $CD8^+$  T cells either increase (12,15) or decrease (16) with age, or do not change at all (17,18).

Comparison of immunnophenotyping data between Asian and non-Asian populations has indicated that the Asian population tend to have lower levels of  $CD3^+$  and  $CD4^+$  T cells and  $CD4^+/CD8^+$  ratio (19).

Comparing our data with previous studies in other regions of Asia, we point out some differences as depicted in Table 5 (20-25).

Comparisons of T and B lymphocytes among different adult population in Iran, Turkey, Kuwait and China did not show any significant difference. We observed higher levels for the mean percentage of CD3<sup>+</sup>, CD4<sup>+</sup> and CD8<sup>+</sup> T cells and lower values for the mean percentage of CD19<sup>+</sup> T cells than the previously mentioned report in Iran (19,20). Another noticeable finding was a lower mean CD4<sup>+</sup>/CD8<sup>+</sup> ratio in our study compared to the pervious study. This was reflected by a higher CD8<sup>+</sup> count in the population of our study. Although our current study showed that the absolute number of CD3<sup>+</sup> T lymphocytes was 1800/mm<sup>3</sup>, which was much higher than the reported values in previous studies, the difference was insignificant. Indeed, the number of B lymphocytes in this study was quite similar to previous studies done in Iran and Kuwait, which were all insignificantly higher than of those studies in Turkey and China. Methodological features, environmental factors (smoking, poor nutrition, infection), and genetics could also contribute to the differences revealed among these populations.

In conclusion, this investigation leads to the establishment of gender-related reference ranges of immunohematological parameters for the Iranian population with 20 to 40 years of age. Since the age of our study population is between 20 to 40 years, which is considered to be a narrow age-range, it would be difficult to come to a definitive conclusion about the impact of age on lymphocyte subsets. Therefore, we recommend recruiting volunteers of a wider age range for future studies.

Population Year of study	CD3 count	CD3(%)	CD4 count	CD4(%)	CD8 count	CD8(%)	CD19 count	CD19(%)	CD56 count	CD56(%)	Ratio
Our study	$1800.87 \pm 471$	$70.9 \pm 7.54$	1039.99 ± 338.02	41.04 ±7.86	783.95 ±234.87	31.11 ± 6.60	328.37±153.17	12.7 ± 4.56	388.62 ±176.17	$15.53 \pm 6.34$	$1.39 \pm 0.48$
Iran 2011(19)	$1504\pm505$	67.66 ± 7.76	827 ± 313	39.22 ± 6.7	522 ± 185	25.42 ± 5.4	332 ± 186	14.41 ± 5.09	NA*	NA*	$1.6 \pm 0.46$
Malay 1993(20)	NA*	69.14±10.30	NA*	$34.14 \pm 8.41$	NA*	36.23 ± 9.96	NA*	11.14 ± 3.62	NA*	NA*	$1.07\pm0.43$
India 2004(21)	NA*	$68.65\pm8.24$	NA*	37.10 ±7.83	NA*	$34.04\pm8.79$	NA*	$14.67 \pm 5.42$	NA*	$12.44 \pm 7.55$	$1.16 \pm 0.41$
China 1996(22)	$1370\pm400$	$69.0 \pm 7.7$	$725 \pm 256$	36.4 ± 7.5	$589\pm205$	$29.7\pm7.2$	221 ± 108	11.1 ± 3.9	$394 \pm 194$	$19.8 \pm 8.1$	$1.0 \pm 0.4$
Turkey 2005 (23)	$1680\pm528$	72.70 ±8.44	$1095 \pm 391$	47.37 ± 9.10	$669 \pm 239$	28.99 ± 5.99	254 ± 122	$10.96 \pm 4.44$	161 ± 92	7.03 ± 3.26	$1.68 \pm 0.43$
Kuwait 2002 (24)	$1770 \pm 470$	NA*	$1050\pm300$	NA*	$730 \pm 220$	NA*	330 ± 140	NA*	320 ± 130	NA*	$1.50 \pm 0.35$

 Table 5. Comparison of T-lymphocyte subsets in different populations.

\*NA= Not available

Nevertheless, these reference ranges are applicable for immunodeficient patients such as HIV-positive patients in the mentioned age range and also individuals who have a high risk of HIV infection.

It should be kept in mind that these results may be reliable for the follow-up of HIVpositive patients as well as for a variety of diseases that involve alterations in lymphocyte sub-populations such as lymphoma, leukemia, etc.

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