Association between *KIR* Genes and Efficacy of Treatment of HBeAg-Positive Chronic Hepatitis B Patients with Entecavir

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ABSTRACT

Background: Entecavir (ETV) is commonly used to treat chronic hepatitis B (CHB) in China. However, certain percentages of e-Antigen (HBeAg) positive CHB patients do not respond to ETV therapy. Objective: To investigate whether the killer immunoglobulin-like receptor (KIR) genes were associated with seroconversion in HBeAg positive CHB responder patients treated with ETV. Methods: Polymerase chain reaction with sequence-specific primers (PCR-SSP) method was performed to genotype KIR genes in 200 healthy controls and 198 HBeAg-positive CHB patients which 59 were defined as the complete response group (CRG) to the treatment with ETV and 139 were defined as null or partial response group (NPRG). Results: The frequencies of KIR2DS2 and KIR2DS3 were significantly higher (P=0.030, OR=1.57, 95%CI=2.36-1.05 and P=0.018, OR=1.773, 95%CI=2.77-1.13, respectively), while, the frequencies of KIR2DL3, KIR2DS1 and KIR3DS1 were significantly lower (P=0.038, OR=0.525, 95%CI=0.96-0.29,and P=0.031, OR=0.640, 95%CI =0.95-0.43, and P=0.035, OR=0.641, 95%CI =0.96-0.43, respectively) in HBeAg-positive CHB patients than those in healthy controls. The frequency of KIR2DS3 gene was significantly higher in NPRG than that in CRG (P=0.018, OR=0.402, 95%CI=0.83-0.20). The frequencies of KIR2DL3 and KIR3DS1 genes were significantly higher in CRG than those in NPRG (P=0.019, OR=3.625, 95%CI=10.83-1.21 and P=0.041, OR=1.949, 95%CI=3.65-1.04, respectively). Conclusion: Patients with KIR2DS3 might have negative responses to anti-HBV therapy with ETV and patients with KIR2DL3 and KIR3DS1 might have advantage in the therapy with ETV.

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Keywords: Entecavir, HBeAg-Positive CHB Patients, KIRGenes

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INTRODUCTION

Hepatitis B virus (HBV) infection is one of the global health concerns affecting about 350 million people (more than 200 million in China) around the world(1). Clinically, HBV infection involves severe consequences such as liver functional failure, fibrosis, cirrhosis, and hepatocellular carcinoma (HCC) since it can survive in the host for the whole life (2). Entecavir (ETV) is the drug of first choice recommendation in chronic hepatitis B (CHB) therapy because of its higher antiviral characteristics and higher genetic barriers than other antiviral agents (3,4). In this regard, the antiviral therapy could ameliorate the progression of liver injury, cirrhosis, and HCC (5-7). The amounts of HBV DNA were less than 300 copies/mL in 80% of HBeAg-positive patients after ETV therapy at 0.5 mg daily through 96 weeks (8). However, there are still some portions of HBeAg-positive CHB patients that do not respond to the ETV therapy, making the molecular mechanism remain largely unknown.

Natural killer (NK) cells play key roles in the control of viral hepatitis and the pathogenesis of liver injury and inflammation (9). The activations of NK cells are of paramount importance in liver inflammation during chronic HBV infection in both HBV transgenic mice and HBV-infected patients (10-12). NK cell activities are regulated by several kinds of regulatory receptors such as the killer immunoglobulinlike receptor (KIR) family (13). KIR gene family makes clusters on human chromosome 19q13.4, which contains 7 inhibitory KIR genes (KIR3DL1-3, KIR2DL1-3, and KIR2DL5), 6 activating KIR genes (KIR3DS1 and KIR2DS1-5), and 1 (KIR2DL4) with both inhibitory and activating characteristics and 2 pseudogenes (KIR2DP1 and KIR3DP1). KIR regulates the activations of NK cells and certain T cells by providing activating or inhibitory signals and possesses the potential for anti-virus applications (14). KIR gene polymorphisms are associated with susceptibility to or protection from infectious diseases such as Treponema pallidum (15), HIV(16), hepatitis C virus (HCV) (17), HBV (18), Ebola virus (19), and Mycobacterium tuberculosis(20). However, whether KIR genes are associated with seroconversion and HBV DNA suppression in HBeAg-positive CHB patients treated with ETV remains unclear.

In this study, we investigated the association of *KIR* genes with seroconversion in HBeAg-positive CHB patients with ETV therapy. For this purpose, we analyzed the frequency of *KIR* genes in 198 ETV-treated HBeAg-positive CHB patients and 200 healthy blood donors in Chinese Han population through polymerase chain reaction with sequence-specific primers (PCR-SSP) method.

MATERIALS AND METHODS

Patients and Controls. Between October 2010 and August 2015 (retrospective type study), a total of 198 consecutive ETV-treated HBeAg-positive CHB patients including 106 men and 92 women with mean age of 40.4 ± 12.6 years were recruited and treated with ETV for 48 weeks. The mean HBV DNA copies were (7.22 ± 1.20) log10 copies/mL in the HBsAg-positive CHB patients. Patients were excluded if they had previous antiviral therapy for HBV, HCV, hepatitis D virus, HIV infections, and liver cirrhosis or HCC.A 48-weeks ETV therapy (0.5 mg daily) was defined as the primary efficacy end point with the changes in seroconversion and HBV DNA suppression.

After the therapy, patients with negative HBeAg and HBV DNA copies less than 300 were defined as complete response group (CRG), those with positive HBeAg and HBV DNA copies more than 300 were defined as null response group (NRG), those with positive HBeAg and HBV DNA copies less than 300 were defined as partial response group (PRG), and the last two situations were together called null or partial response group (NPRG). Meanwhile, 200 regular blood donors were recruited as healthy controls (no HIV, HCV,HBV, syphilis infections, and normal alanine aminotransferase (ALT) level) including 102 males and 98 females with mean age of 43.5±6.1 years. The patients and healthy blood donors consent to participate in the study. We had the ethical statement (NO.2010017) approved by the Ethics Committee of the Blood Center of Shandong Province.

Laboratory Methods.HBV DNA copies in blood serum were examined by real-time PCR method with a linear dynamic detection range from 75 copies/mL to 5×10^9 copies/mL (Abbott Laboratories, Chicago, IL, USA). The characteristics of ALT HBeAg, HBsAb, HBeAg, HBeAb, and HBcAb in patients' serum were measured by enzyme immunoassays method (lower limit of detection (LLOD):0.05 IU/mL, Abbott Laboratories, Chicago, IL, USA).

KIR Gene Distributions. The 16 KIR genes were examined by PCR-SSP method (15) in recruited subjects. When the DNA from frozen peripheral blood mononuclear cells was extracted (EZ Bead System-32 DNA workstation, Texas BioGene Inc., USA), the PCR-SSP typing of KIR genes were done as soon as possible to avoid false-negative results due to the longer KIR-specific amplicons. The human growth hormone gene (15) was used as positive control while water was as a negative control. All primers for the PCR-SSP were bought from BOYA. Bio Co., Ltd., Shanghai. The 10 µl volume PCR system contained 20-50 ng DNA ,0.2 mM dNTP, 0.5U Taq DNA polymerase (Promega), 0.4 µM primers (except for KIR2DS1, 0.8 µM), and 1X PCR buffer. The conditions of PCR amplification in a 9700 thermal cycler (PerkinElmer, Waltham, MA, USA) were as follows: initial denaturing at 94 for 4 min, followed by 30 cycles of 94 for 30sec, 72 for 90sec, plus a final extension at 72°C for 10 min. The for 30sec, 65 annealing temperatures of KIR2DS2, KIR2DS3, and KIR2DS5 were 63 while KIR2DS4 was 61 . About 1-2% fluorescence-dyed agarose gels were used to analyze the PCR products (Gene Genius Bio Imaging System, Syngene Ltd., UK).

Statistical Analysis. *KIR* gene frequencies were analyzed according to the description given in (15). The direct counting method was used for the analysis of phenotypic frequencies (PF). The significance difference analysis of *KIR* genes' frequencies was performed by Yates' correction analysis ($P \le 0.05$). The odds ratio (OR) and relative risk (RR) were analyzed using the SPSS13.0 software package.

RESULTS

Patients' Characteristics.

All the 198 HBeAg-positive CHB patients showed different outcomes after 48 weeks ETV therapy. Of this population, 29.8% of the patients were classified as CRG while others were classified as NPRG including NRG (10.1%) and PRG (60.1%), Table 1. The patients with reduced HBV DNA (<300 copies/mL) were 89.9%, including CRG and PRG. The ALT levels showed significant differences between CRG and NRG (P=0.00016).

	n (%)	ALT (U/L)	HBV DNA (copies/mL)	HBsAg	HbsAb	HBeAg	HBeAb	HBcAb
CRG	59(29.80)	30.21±7.25	<300	+	-	-	+	+
NRG	20(10.10)	132.67±10.58	>300	+	-	+	-	+
PRG	119(60.10)	48.29±9.17	<300	+	-	+	-	+

Table1. The characteristics of HBeAg-positive CHB patients after 48-weeks-treatment with ETV.

CHB: Chronic Hepatitis B; ETV: Entecavir; n: numbers of patients; CRG: Complete Response Group Treated with ETV; NRG: null response group treated with ETV, PRG: partial response group treated with ETV; ALT: alanine aminotransferas;+: positive; -: negative.

KIR Gene Frequencies in Healthy Controls and HBeAg-Positive CHB Patients.

A total of 16 *KIR* genes were detected in healthy controls and HBeAg-positive CHB patients(Table 2) while *KIR2DL4*, *KIR3DL2*, *KIR3DL3*, and *KIR3DP1* were found in all subjects as the framework genes. The inhibitory *KIR* genes presented commonly higher frequencies than activating types in the two groups except for *KIR2DL2* and *KIR2DL5*. The frequencies of activating *KIR2DS2* and *KIR2DS3* increased significantly in HBeAg-positive CHB patients than those in controls (P=0.030, OR=1.577, and 95%CI=2.36-1.05; P=0.018, OR=1.773, and 95%CI=2.77-1.13, respectively) while the frequencies of *KIR2DL3*, *KIR2DS1*, and *KIR3DS1* were significantly decreased in CHB patients (P=0.038, OR=0.525, and 95%CI=0.96-0.29;P=0.031, OR=0.640, and 95%CI=0.95-0.43; and P=0.035, OR=0.641, 95%CI =0.96-0.43, respectively). There was no statistically significant difference in distributions of other *KIR* genes in the healthy controls and HBeAg-positive CHB patients.

KIR Gene Frequencies in CRG and NPRG HBeAg-Positive CHB Patients with ETV Therapy.

After treatment with ETV for 48 weeks, 59 of 198 HBeAg-positive CHB patients were CRG responders and 139 were NPRG responders. The distributions of *KIR* genes between the CRG and NPRG patients are shown in Table 3.The results show a significant increase in the frequency of *KIR2DS3* significantly (P=0.018, OR=0.402, and 95%CI=0.83-0.20) and a significant decrease in the frequencies of *KIR2DL3* and *KIR3DS1* (P=0.019, OR=3.625, and 95%CI=10.83-1.21; P=0.041, OR=1.949, and 95%CI=3.65-1.04, respectively) in NPRG patients compared with CRG patients. There were no significantly different distributions of other *KIR* genes in the two groups.

KIR genes	Healthy controls $(n=200)$		CHB patients (n=198)		P-value	OR	95%CI
	+	gf(%)	+	gf(%)	-		
2DL1	195	97.50	188	94.95	0.187	0.482	1.44-0.16
2DL2	53	26.50	46	23.23	0.456	0.839	1.32-0.53
2DL3	181	90.50	165	83.33	0.038*	0.525	0.96-0.29
2DL4	200	100.00	198	100.00	-	-	-
2DL5	64	32.00	76	38.38	0.185	1.324	2.00-0.88
3DL1	177	88.50	172	86.87	0.615	0.860	1.56-0.47
3DL2	200	100.00	198	100.00	-	-	-
3DL3	200	100.00	198	100.00	-	-	-
2DS1	106	53.00	83	41.92	0.031*	0.640	0.95-0.43
2DS2	71	35.50	92	46.46	0.030 *	1.577	2.36-1.05
2DS3	44	22.00	66	33.33	0.018*	1.773	2.77-1.13
2DS4	154	77.00	165	83.33	0.122	1.494	2.46-0.91
2DS5	63	31.50	56	28.28	0.476	0.858	1.32-0.56
3DS1	91	45.50	69	34.85	0.035*	0.641	0.96-0.43
2DP1	193	96.50	185	93.43	0.156	0.516	1.32-0.20
3DP1	200	100.00	198	100.00	-	-	-

Table 2. The frequencies of *KIR* genotypes in healthy controls and HBe Agpositive CHB patients.

+: numbers of each genotype; gf: genotype frequencies; *: indicates statistical significance (P < 0.05).

KIR genes	CRG (n=59)		NPRG (n=139)		<i>P</i> -value	OR	95%CI
	+	gf(%)	+	gf(%)	-		
2DL1	57	96.61	131	94.24	0.492	1.740	8.45-0.36
2DL2	17	28.81	29	20.86	0.231	1.535	3.08-0.77
2DL3	55	93.22	110	79.14	0.019*	3.625	10.83-1.21
2DL4	59	100.00	139	100.00	-	-	-
2DL5	27	45.76	49	35.25	0.169	1.550	2.88-0.83
3DL1	49	83.05	123	88.49	0.308	0.637	1.50-0.27
3DL2	59	100.00	139	100.00	-	-	-
3DL3	59	100.00	139	100.00	-	-	-
2DS1	25	42.37	58	41.73	0.939	1.027	1.90-0.55
2DS2	26	44.07	66	47.48	0.667	0.871	1.61-0.47
2DS3	12	20.34	52	38.85	0.018*	0.402	0.83-0.20
2DS4	46	77.97	119	85.61	0.192	0.595	1.29-0.27
2DS5	16	27.12	40	28.78	0.820	0.921	1.82-0.47
3DS1	27	45.76	42	30.22	0.041*	1.949	3.65-1.04
2DP1	53	89.83	132	94.96	0.189	0.468	1.46-0.15
3DP1	59	100.00	139	100.00	-	-	-

Table 3. The distributions of the frequencies of *KIR* genes in CRG and NPRG patients.

CRG: complete response group treated with ETV; NPRG:null or partial response group treated with ETV; +: numbers of each genotype; gf: genotype frequencies; *: indicates statistical significance (P < 0.05).

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DISCUSSION

Through anti-HBV treatment, the progression of chronic liver diseases such as cirrhosis and HCC was ameliorated and HBV DNA replication was suppressed; otherwise, their HBV DNA levels increased gradually (21,22). Therefore, the reduction levels of HBV DNA copies are defined as a critically clinical therapeutic aim for CHB patients. Currently, ETV is one of the most common anti-HBV drugs used in China. During the therapy of HBV infection in the CHB patients, HBeAg seroconversion serves as an important landmark in the control of HBV infectious progression. Earlier HBeAg seroconversion with low HBV DNA copies usually confers a favorable clinical outcome. NK cells possess a critical role in anti-virus immunity. In this regard, ETV promotes NK cells functions (23,24). The numbers of NK cells increased in CHB patients treated with ETV(23). ETV therapy for HBeAg-positive CHB patients declined the number of HBV DNA and ALT levels became normal; besides, it led to the recovery of NK cellmediated immunity (24). KIRs regulate the activations of NK cells(14). Therefore, the HBeAg-positive CHB patients undergoing 48 weeks ETV therapy were chosen to determine whether the suppression of HBV DNA replication and HBeAg seroconversion was associated with KIR genes and ETV treatment. To our knowledge, there has been no report on the effects of KIR genes on efficacies of HBeAg-positive CHB patients with ETV therapy.

ETV was effective to reduce HBV DNA (<300 copies/mL) in 89.9% and decrease HBeAg seroconversion in 29.8% of the HBeAg-positive CHB patients in this study (Table 1); in line with the results of previous reports (22,25). The amount of HBV DNA is one of the crucial components for evaluating the efficacies of anti-HBV treatment in the CHB patients. The efficacies of ETV therapy might be due to its potent functions in suppression of HBV replication. Further, these results indicate that there were still some patients with poor responses to ETV treatment in the early stage.

As the immune system plays important roles in the amelioration of HBV-infection, it is a promising area to investigate predictive markers of response to ETV therapy. The activities of NK cells and certain T cells has been shown to be important functions in reducing amounts of HBV and in controlling the development processes of liver injury(9,26-28). The KIR genes can regulate the activations of NK cells. Moreover, certain T cells responded to many kinds of microbial pathogens (14), suggesting that KIR gene diversities may affect susceptibilities to or protection from variable infections. The previous discoveries indicated that KIR genes were associated with infectious diseases caused by Treponema pallidum (15), HIV (16), HCV (17), HBV (18), Ebola virus(19), *Mycobacterium* tuberculosis(20), malaria(29), and *Mycobacterium* leprae(30)infection, indicating that different KIR genes might be of importance in the susceptibility or clearance to different infections.

The results of the present study showed that the *KIR2DS2* and *KIR2DS 3* were associated with susceptibility of, whereas the *KIR2DS1,KIR3DS1*,and *KIR2DL3* might be protective against HBV infection, suggesting that different *KIR* genes might show the variable immune responses to HBV infection. These results were partly different from a previous report(18), which showed *KIR2DL5* was protective, but not *KIR2DL3*.We considered that the main possible reasons were the different patient groups, which one was CHB patient group (18) and the other was HBeAg-positive CHB patient group in our study. It is of note that the HBeAg-positive CHB patients with *KIR* genes showed significantly different responses to ETV therapy. The patients with

KIR3DS1 and KIR2DL3 were CRG that facilitated the seroconversion and reduction of the amount of HBV, whereas the patients with the KIR2DS3 were NPRG. This study was the first to research KIR genes associated with efficacies of HBeAg-positive CHB patients with ETV therapy. These findings suggested that different KIR genes might use combinations of synergistic receptors to activate NK cells for benefit of HBV clearance. A recent study showed that KIRs expressed on NK cell surface play an important role in regulating immune responses through the transducing or activating inhibitory signals(31). Previous investigations have shown that the frequency of KIR2DS3 was significantly increased in syphilis (15), hepatitis C (17), CHB (18), and hemorrhagic fever (19) patients compared to those in the respective controls. It was suggested that KIR2DS3 might involve in the physiopathological process by excessively destroying host's cells due to its favoring role in the host's NK cells activation, supporting a result that activating KIR has an association with disease susceptibility to infections (19). Interestingly, the variations of KIR2DS3 and IL28B were strongly related with clinical responses to both pegylated-IFN and ribavirin treatments in an HIV-1/HCV co-infected patient. This result demonstrates that testing for host certain genetic genes would be useful for precision medicine in infected patients. Besides, it provides further evidence that the innate immune system is of importance in protection from HIV-1/HCV infections(32). The KIR3DS1 with its HLA-B ligand gene was related with a slower development process for AIDS in HIV-infected individuals, suggesting that KIR3DS1 with and its ligand were involved protective responses of NK cells after HIV-1 infection (16). Moreover, KIR3DS1 homozygotes showed the significantly slower progression of HIV infection than KIR3DL1/S1 heterozygotes (33).HIV infection promotes the activations of NK cell receptors and its ligands, which in turn stimulates NK cells activities such as secreting chemokines to protect from HIV(34,35). The frequency of KIR2DL3 was lower in CHB than in subjects with resolved HBV infection, which indicates a protective role of KIR2DL3(36). This result is in line with our findings. The mechanism responsible for the protective role of KIR2DL3 is unclear, as this gene codes for an inhibitory receptor. Lisovsky et al. showed that KIR2DL3 ⁺ NK cells were mediators of HIV-specific responses (37). The interaction of the KIR2DL3-HLA-C1 was associated with resolved infection in HCV (38), Treponema pallidum(15), and HBV(39), suggesting a possible generalizability of the protective role of KIR2DL3 to different infections. KIR2DL3 has a weaker affinity to HLA-C1 than KIR2DL2. Therefore, it was suggested that this weaker interaction between the inhibitory receptor and its ligand (KIR2DL3-HLA-C1) would have protective properties because of the more facile overriding through activating signals (38). Those studies suggested that virus infection was associated with the host's KIR gene distributions and that NK cells might play a critical role in protection from or susceptible to the infectious diseases. The biological functions of KIR gene distributions in HBeAg-positive CHB pathogenesis, however, are still unclear and need further research.

In conclusion, the certain *KIR* genes might play a critical role in the sustain infection or cure of CHB patients undergoing ETV therapy. Our findings might be useful for predicting the precision medicine in HBeAg-positive CHB patients with ETV therapy. Future studies on the mechanism underlying these genetic associations might provide insights for new therapeutic strategies in HBV infected patients.

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REFERENCES

- 1. Lu Z, Zhang B, Chen S, Gai Z, Feng Z, Liu X, et al. Association of KIR genotypes and haplotypes with susceptibility to chronic hepatitis B virus infection in Chinese Han population. Cell Mol Immunol. 2008; 5:457-63.
- Huang YJ, Chang CS, Peng YC, Yeh HZ, Yang SS. On-treatment HBV DNA dynamics predict virological breakthrough in entecavir-treated HBeAg-positive chronic hepatitis B. PLoS ONE. 2017; 12: e0174046.
- 3. Leung N, Peng CY, Hann HW, Sollano J, Lao-Tan J, Hsu CW, et al. Early hepatitis B virus DNA reduction in hepatitis B e antigen-positive patients with chronic hepatitis B: A randomized international study of entecavir versus adefovir. Hepatology.2009; 49:72-9.
- 4. European Association for the study of the Liver. EASL clinical practice guidelines: management of chronic hepatitis B virus infection. J Hepatol.2012; 57:167-85.
- 5. Jang JW, Choi JY, Kim YS, Woo HY, Choi SK, Lee CH, et al. Long-term effect of antiviral therapy on disease course after decompensation in patients with hepatitis B virus-related cirrhosis. Hepatology. 2015; 61:1809-20.
- 6. Papatheodoridis GV, Chan HL, Hansen BE, Janssen HL, Lampertico P. Risk of hepatocellular carcinoma in chronic hepatitis B: assessment and modification with current antiviral therapy. J Hepatol. 2015; 62:956-67.
- 7. Wu CY, Chen YJ, Ho HJ, Hsu YC, Kuo KN, Wu MS, et al. Association between nucleoside analogues and risk of hepatitis B virus-related hepatocellular carcinoma recurrence following liver resection. JAMA. 2012; 308:1906-14.
- 8. Gish RG, Lok AS, Chang TT, de Man RA, Gadano A, Sollano J, et al. Entecavir therapy for up to 96 weeks in patients with HBeAg-positive chronic hepatitis B. Gastroenterology. 2007; 133:1437-44.
- 9. Tian Z, Chen Y, Gao B. Natural killer cells in liver disease. Hepatology. 2013; 57:1654-62.
- Dunn C, Brunetto M, Reynolds G, Christophides T, Kennedy PT, Lampertico P, et al. Cytokines induced during chronic hepatitis B virus infection promote a pathway for NK cell-mediated liver damage. J Exp Med. 2007; 204:667-80.
- 11. Chen Y, Sun R, Jiang W, Wei H, Tian Z. Liver-specific HBsAg transgenic mice are oversensitive to Poly(I:C)-induced liver injury in NK cell- and IFN-gamma-dependent manner. J Hepatol. 2007; 47:183-90.
- 12. Trachtenberg E, Vinson M, Hayes E, Hsu YM, Houtchens K, Erlich H, et al. HLA class I (A,B, C) and class II (DRB1, DQA1, DQB1, DPB1) alleles and haplotypes in the Han from southern China. Tissue Antigens. 2007;70:455-63.
- 13. Parham P. MHC class I molecules and KIRs in human history, health and survival. Nat Rev Immunol. 2005; 5:201-14.
- 14. Carrington M, Martin MP. The impact of variation at the KIR gene cluster on human disease. Curr Top Microbiol Immunol. 2006; 298:225-57.
- 15. Zhuang YL, Ren GJ, Tian KL, Li XY, Zhu YB, Liu JL, et al. Human leukocyte antigen-C and killer cell immunoglobulin-like receptor gene polymorphisms among patients with syphilis in a Chinese Han population. APMIS. 2012; 120:828-35.
- 16. Martin MP, Gao X, Lee JH, Nelson GW, Detels R, Goedert JJ, et al. Epistatic interaction between KIR3DS1 and HLA-B delays the progression to AIDS. Nat Genet. 2002; 31: 429-34.
- 17. Dring MM, Morrison MH, McSharry BP, Guinan KJ, Hagan R. Irish HCV Research Consortium, O'Farrelly C and Gardiner CM. Gardiner, Innate immune genes synergize to predict

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increased risk of chronic disease in hepatitis C virus infection. Proc Natl Acad Sci U S A. 2011; 108:5736-41.

- Lu Z, Jiao Y, Feng Z, Dong Z, hang B, Zhao Y. Polymorphisms of killer cell immunoglobulinlike receptor gene: possible association with susceptibility to or clearance of hepatitis B virus infection in Chinese Han population. CMJ. 2007;48:800-6.
- 19. Wauquier N, Padilla C, Becquart P, Leroy E, Vieillard V. Association of KIR2DS1 and KIR2DS3 with fatal outcome in Ebola virus infection. Immunogenetics. 2010;62:767-71.
- 20. Méndez A, Granda H, Meenagh A, Contreras S, Zavaleta R, Mendoza MF, et al. Study of KIR genes in tuberculosis patients. Tissue Antigens. 2006;68:386-9.
- Lai CL, Shouval D, Lok AS, Chang TT, Cheinquer H, Goodman Z, et al. Entecavir versus lamivudine for patients with HBeAg-negative chronic hepatitis B. N Engl J Med. 2006;354:1011-20.
- Chang TT, Gish RG, De Man R, Gadano A, Sollano J, Chao YC, et al. A comparison of entecavir and lamivudine for HBeAg-positive chronic hepatitis B. N Engl J Med. 2006;354:1001-10.
- 23. Zhang L, Wang Q, Zhao P, Hu X, Jiang Y. Effects of entecavir on peripheral blood lymphocyte profiles in chronic hepatitis B patients with suboptimal responses to adefovir. Clin Exp Pharmacol Physiol. 2014;41:514-23.
- Zhao PW, Jia FY, Shan YX, Ji HF, Feng JY, Niu JQ, et al. Downregulation and altered function of natural killer cells in hepatitis B virus patients treated with entecavir. Clin Exp Pharmacol Physiol. 2013;40:190-6.
- Xu JH, Wang S, Xu ZN, Yu YY, Si CW, Zeng Z, et al. Entecavir maleate versus entecavir in Chinese chronic hepatitis B predominantly genotype B or C: results at week 144. J Viral Hepat. 2017; 24:877-84.
- Maini MK, Boni C, Ogg GS, King AS, Reignat S, Lee CK, et al. Direct ex vivo analysis of hepatitis B virus specific CD8(+) T cells associated with the control of infection. Gastroenterology. 1999;117:1386-96.
- 27. Stelma F, De Niet A, Tempelmans Plat-Sinnige MJ, Jansen L, Takkenberg RB, Reesink HW, et al. Natural Killer Cell Characteristics in Patients With Chronic Hepatitis B Virus (HBV) Infection Are Associated With HBV Surface Antigen Clearance After Combination Treatment With Pegylated Interferon Alfa-2a and Adefovir. J Infect Dis. 2015; 212:1042-51.
- Micco L, Peppa D, Loggi E, Schurich A, Jefferson L, Cursaro C, et al. Differential boosting of innate and adaptive antiviral responses during pegylated-interferon-alpha therapy of chronic hepatitis B. J Hepatol. 2013; 58:225-33.
- 29. Artavanis-Tsakonas K, Eleme K, McQueen KL, Cheng NW, Parham P, Davis DM, et al. Activation of a subset of human NK cells upon contact with Plasmodium falciparum-infected erythrocytes. J Immunol. 2003;171:5396-405.
- Franceschi DSA, Mazini PS, Rudnick CCC, Sell AM, Tsuneto LT, De Melo FC, et al. Association between killer-cell immunoglobulin-like receptor genotypes and leprosy in Brazil. Tissue Antigens. 2008;72:478-82.
- Khakoo SI, Carrington M. KIR and disease: a model system or system of models? Immunol Rev. 2006; 214:186-201.
- 32. Keane C, O'Shea D, Reiberger T, Peck-Radosavljevic M, Farrell G, Bergin C, et al. Variation in both IL28B and KIR2DS3 genes influence pegylated interferon and ribavirin hepatitis C treatment outcome in HIV-1 co-infection. PLoS One. 2013; 8:e66831.
- Tallon BJ, Bruneau J, Tsoukas CM, Routy JP, Kiani Z, Tan X, et al. Time to seroconversion in HIV-exposed subjects carrying protective versus non protective KIR3DS1/L1 and HLA-B genotypes. PloS one. 2014; 9:e110480.
- 34. Fauriat C, Long EO, Ljunggren HG, Bryceson YT. Regulation of human NK-cell cytokine and chemokine production by target cell recognition. Blood. 2010;115:2167-76.
- Song R, Lisovsky I, Lebouché B, Routy JP, Bruneau J, Bernard NF. HIV protective KIR3DL1/S1-HLA-B genotypes influence NK cell-mediated inhibition of HIV replication in autologous CD4 targets. PLoS Pathog. 2014;10:e1003867.
- Di Bona D, Aiello A, Colomba C, Bilancia M, Accardi G, Rubino R, et al. KIR2DL3 and the KIR ligand groups HLA-A-Bw4 and HLA-C2 predict the outcome of hepatitis B virus infection. J Viral Hepat. 2017;24:768-75.

- 37. Lisovsky I, Isitman G, Tremblay-McLean A, Song R, DaFonseca S, Lebouché B, et al. The differential impact of natural killer (NK) cell education via KIR2DL3 and KIR3DL1 on CCL4 secretion in the context of in-vitro HIV infection. Clin Exp Immunol. 2016;186:336-46.
- 38. Khakoo SI, Thio CL, Martin MP, Brooks CR, Gao X, Astemborski J, et al. HLA and NK cell inhibitory receptor genes in resolving hepatitis C virus infection. Science. 2004; 305:872-4.
- Gao X, Jiao Y, Wang L, Liu X, Sun W, Cui B, et al. Inhibitory KIR and specific HLA-C gene combinations confer susceptibility to or protection against chronic hepatitis B. Clin Immunol. 2010; 137:139-46.